Some amino sugars structurally related to 6-

Carbohydr. Res. 2002, 337, 663

deoxymannojirimycin precursors prepared from methyl 6-deoxy-2,3-O-isopropylidene- α -D-lyxo-hexofuranosid-5-ulose and methyl 2,3-O-isopropylidene- α -D-lyxo-pentodialdo-1,4-furanoside

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R = Me, H $X, Y = CN, NHR^{1}, COOEt$

An access to various sulfation patterns in dermatan sulfate: chemical syntheses of sulfoforms of trisaccharide methyl glycosides

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 $\begin{array}{c} \text{NaSO}_{3}\text{O} \\ \text{OH} \\ \text{OH} \\ \text{OH} \\ \text{OH} \\ \text{OH} \\ \text{OH} \\ \text{ONO}_{3}\text{Na} \\ \text{NaSO}_{3}\text{O} \\ \text{OH} \\ \text$

Synthesis of oxidized methyl 4-O-methyl- β -D-glucopyranoside and methyl β -D-glucopyranosyl- $(1 \rightarrow 4)$ - β -D-glucopyranoside derivatives as substrates for fluorescence labeling reactions

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The chemical synthesis, oxidation, and fluorescence labelling of mono- and disaccharide derivatives serving as model compounds for oxidized celluloses is described.

MeO OH OH OH OH

Regulation of fructosyltransferase activity by carbohydrates, in solution and immobilized on hydroxyapatite surfaces

Doron Steinberg, Ramona Rozen, Moshe Bromshteym, Batia Zaks, Itzhak Gedalia, Gilad Bachrach Department of Oral Biology, Faculty of Dentistry, Hebrew University-Hadassah, PO Box 12272, Jerusalem 91120, Israel

We tested the effect of several carbohydrates on the activity of cell-free fructosyltransferases (FTF) in solution and immobilized onto hydroxyapatite (HA) and found an inhibitory dose-dependent effect of glucose on FTF activity, both on the surface and in solution.

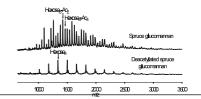
Carbohydr. Res. 2002, 337, 711

Characterization of water-soluble hemicelluloses from spruce and aspen employing SEC/MALDI mass spectroscopy

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Water-soluble hemicelluloses were isolated from spruce and aspen employing microwave treatment. The degree of substitution with acetyl moieties (DS) of the acetylated hemicelluloses was calculated on the basis of the MALDI-MS spectra obtained prior to and following deacetylation.



Structure of a fucoidan from the brown seaweed Fucus evanescens C.Ag.

Carbohydr. Res. 2002, 337,

Maria I. Bilan, Alexey A. Grachev, Nadezhda E. Ustuzhanina, Alexander S. Shashkov, Nikolay E. Nifantiev, Anatolii I. Usov

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Extracellular polysaccharides of a bacterium associated with a fungal canker disease of Eucalyptus sp

Carbohydr. Res. 2002, 337, 731

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Extracellular polysaccharides (EPSs) produced by an Erwinia sp associated with a fungal canker disease of Eucalyptus were fractionated into one polysaccharide that was identified with that produced by Erwinia chrysanthemi strains SR260, Ech1, and Ech9, and the other distinctively different from any other EPS produced by E. chrysanthemi strains so far studied. Their structures were determined using a combination of chemical and physical techniques including methylation analysis, low pressure gel-filtration and anion-exchange chromatographies, high-pH anion-exchange chromatography, mass spectrometry and 1D and 2D 1H NMR spectroscopy. The new polysaccharide, identified as EPS Taranera, has the following structure:

$$\rightarrow 3) - \alpha \text{-L-Rhap-}(1 \rightarrow 3) - \alpha \text{-L-Rhap-}(1 \rightarrow 2) - \alpha \text{-L-Rhap-}(1 \rightarrow 3) - \beta \text{-D-Galp}(1 \rightarrow 2) \\ \uparrow \\ \beta \text{-D-Py(4,6)Gicp-}(1 \rightarrow 2) - \beta \text{-D-GilcpA}$$

The molecular weights of the polysaccharides range from $3.2-6.2\times10^5$ and their hydrodynamic properties are those of polydisperse, polyanionic biopolymers with pseudoplastic, non-thixotropic flow characteristics in aqueous solutions.

Carbohydr. Res. 2002, 337, 743 Structural and physicochemical characterization of the inclusion complexes of cyclomaltooligosaccharides (cyclodextrins) with melatonin

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 $n = 6 \alpha$ -CD $n = 7 \beta$ -CD

5-Azido neuraminic acid thioglycoside as sialylation donor

Carbohydr. Res. 2002, 337, 755

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Production of N-acetyl-D-glucosamine from α-chitin by crude enzymes from Aeromonas hydrophila H-2330

Carbohydr. Res. 2002, 337, 761

Oligosaccharides

Exo-chitinase

GlcNAc

(69.4 mU/mg)

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Sakyo-ku, Kyoto 606-8585, Japan
Chitin
□ □ Endo-chitinase
(26.9 mU/mg)

Efficient (yield = 77%) production of GlcNAc was achieved from flake type of α -chitin by crude enzymes derived from *Aeromonas hydrophila* H-2330.

Can the stereochemical outcome of glycosylation reactions Carbohydr. Res. 2002, 337, 765 be controlled by the conformational preferences of the glycosyl donor?

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